



Uso y rendimiento de NGS en diagnóstico prenatal

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Introducción

Anomalías congénitas: 3% de ecografías

Cariotipo, QF-PCR y CMA identifican 40% de anomalías con origen genético

Sin embargo, 70 - 80% de fetos con cariotipo normal tienen CMA normal (Hilman, 2013) ->
¿enfermedades monogénicas?

Shaffer LG et al. Detection rates of clinically significant genomic alterations by microarray analysis for specific anomalies detected by ultrasound. Prenat Diagn. 2012;32:986–95.

Hillman SC et al. Use of prenatal chromosomal microarray: prospective cohort study and systematic review and meta-analysis. Ultrasound Obstet Gynecol. 2013;41:610–20.

Método de elección para enfermedades monogénicas es **secuenciación masiva en paralelo**:

- Paneles (dirigidos a motivo de indicación)
- Secuenciación de exoma (clínico o completo)
- Secuenciación de genoma completo

Indicaciones

- Anormalidad(es) ecográficas **suggerentes de origen monogénico** (al menos 20% de rendimiento esperado*)
- Anomalías múltiples con **cariograma/CMA negativos**
- **Agregación familiar** de fenotipo (feto previo con anomalías o hijo previo afectado)

* Se pueden considerar con rendimientos entre 10 y 20%

Table 1 Clinical indications and estimated diagnostic yield for prenatal NGS (ordered from higher to lower diagnostic yield)

Clinical indication	Diagnostic yield of NGS (%)	Reference
Bilateral hyperechogenic, dysplastic or polycystic kidneys	64	22
Skeletal dysplasia	53	23
Recurrent anomaly	40	24
Fetal akinesia deformation sequence	37	23
Craniosynostosis	38	22
Multiple anomalies involving various systems	33	25
Central nervous system anomalies (except single anomalies)	32	26
Non-isolated nuchal translucency	26	23
Non-immune hydrops fetalis	22	23
NGS, next-generation sequencing.		

Table 2 Anomalies with a diagnostic yield of >10% may also be considered

Clinical indication	Diagnostic yield of NGS (%)	Reference
Single CNS anomaly	16	26
Severe early onset fetal growth restriction	12	27
Isolated heart defect	11	23
CNS, central nervous system; NGS, next-generation sequencing.		

Guidelines for NGS procedures applied to prenatal diagnosis by the Spanish Society of Gynecology and Obstetrics and the Spanish Association of Prenatal Diagnosis

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Although we have provided explanations for different NGS diagnostic approaches (panels, ES, WGS), we strongly recommend the use of ES in prenatal diagnosis for the reasons previously detailed. Therefore, all considerations regarding clinical considerations, reported findings and genetic counselling will be based on the use of ES.

Trio ES (parents and fetus) is recommended in prenatal diagnosis because it allows for filtering out many non-informative variants and it speeds up response time.^{18 19}

Recomendaciones actuales: **preferir exoma completo** por sobre otras alternativas (idealmente exoma trío)

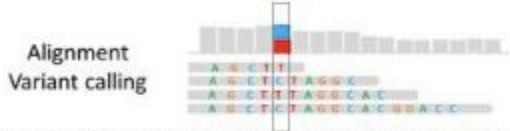
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Consideraciones técnicas



Workflow Step	Consideration
1. Sample processing and library preparation 	DNA quality and Quantity Turnaround Time Maternal Cell Contamination
2. Sequencing and primary bioinformatics analysis 	Variant type: SNVs and indels Allele fractions: CPM, MCC, twins effects
3. Variant filtration  General population Disease databases Mode of inheritance Phenotype?	Lack of prenatal phenotyping Absence of genotype-phenotype information "De novo" filters and MCC
4. Interpretation and reporting  Allele frequency Gene-disease association Functional and segregation data Clinical information	Lack of prenatal phenotyping Absence of genotype-phenotype information Variants of Uncertain Clinical Significance Primary versus secondary findings Implications to other family members
5. Counseling  Pregnancy outcomes Intervention options Uncertainties	Penetrance Age of onset Ethical issues

Método en general es similar a postnatal, pero con algunas consideraciones

Preparación de librería

Similar a cualquier muestra postnatal si la calidad es adecuada

- Fragmentación de ADN genómico (ultrasonificación o procesos enzimáticos), luego ligación de adaptadores

Kits **hasta 50 ng de ADN input**

NGS en muestra de líquido amniótico

Líquido amniótico contiene poblaciones heterogéneas de células derivadas del feto

A diferencia de BVC, tiene **menor tasa de contaminación materna y no tiene mosaicismo placentario confinado**

NGS en muestra de líquido amniótico

Contaminación por ADN materno estimada en **0.3 - 0.7%** del total de muestras

Se cree que provendría de **células de piel materna**

- Se sugiere descartar primeros mL de muestra

Cultivo podría probabilidad de contaminación
(sin embargo, no se hace de rutina)

Mosaicismo en BVC

- Mosaicismo en 1 - 2.5% de BVC por muestra materna
- Cultivar BVC no mejora outcomes y aumenta tiempo de proceso
- Disección cuidadosa de BVC mejora resultados
 - De todas maneras, debiera evaluarse con SNPs o STRs entre sangre materna y muestra
- En caso de detectarse, se recomienda estudio en líquido amniótico

Otras limitaciones

NGS detecta indels hasta 25 - 50 bp de manera confiable

- No confiable clínicamente para CNVs a nivel de exón, expansión de repeticiones o rearrreglos estructurales
- WGS es más sensible que WES para detección de CNVs y rearrreglos

Interpretación limitada a regiones codificantes +/- 2-20 bp de límites codificantes

Detección de variantes missense, indels y CNVs generalmente > 30 kb es diagnóstica

Otras limitaciones

Asociaciones fenotipo-genotipo prenatal están mucho menos descritas y son variables en el tiempo

- Por ende, es más difícil determinar patogenicidad de variantes

Evolving fetal phenotypes and clinical impact of progressive prenatal exome sequencing pathways: cohort study

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D Williams³, M D Kilby^{5 6}



En fetos con variante patogénica causal de fenotipo y que el embarazo alcanzó el tercer trimestre, **se encontró anomalías adicionales en 73.3% de casos**

- Hidrops (27%), artrogriposis (27%) y anomalías cerebrales (18.2%)

Considerations for using prenatal NGS diagnostic tests

- ▶ Prenatal phenotypic relation to most genetic conditions have not been extensively reported. The application of prenatal NGS diagnostic tests will improve our knowledge in prenatal genotype-phenotype conditions (phenotype expansion).¹⁶ It is crucial to take this into account when analysing the variants identified as potentially causatives.
- ▶ The gestational age at which NGS diagnostic tests are performed should be considered, since fetal phenotype might evolve over time. If new clinical information becomes available, either during pregnancy, postnatally or through necropsy findings after termination of pregnancy or fetal demise, exome reanalysis should be considered.

Table 2. Revised Karyotypes from Whole-Genome Sequencing

Subject ID	Clinical Interpretation	Revised Karyotype from Sequencing	Diagnosis
1	46,XY,t(9;16)(q22;p11)	46,XY,t(9;16)(q22.33;p12.1)	Autism spectrum disorder
2	46,XY,t(3;6)(q26.2;q16.2)	46,XY,t(3;6)(q26.32;q16.3)	Autism spectrum disorder
3 ^a	46,XX,inv(5)(p12q13.1)	46,XX,inv(5)(p14.2q14.3)	Global developmental delay, hypotonia, seizures
4	46,XY,t(6;9)(q16.2;q13)	46,XY,inv(6)(q16.1q16.1)t(6;9)(q16.1;q21.3)	Autism spectrum disorder
5a, b	46,XY,t(3;18)(q13.3;q21.3)	46,XY,t(3;18)(q13.32;q21.2)	Global developmental delay, multiple congenital anomalies

Karyotype analyses for each subject included in whole-genome sequencing. Karyotyping was performed at various sites, including referring clinics. In all subjects, a revision to the clinical interpretation was required after sequencing. Subjects included in the CapBP experiment were required to have cytogenetic analyses that previously localized the breakpoint to less than one megabase, so further revision was not necessary.

^a Subject is DGAP218, see Developmental Genome Anatomy Project ([Web Resources](#)).

Rearreglos son detectables por WGS. Sin embargo, sin validación clínica entre laboratorios



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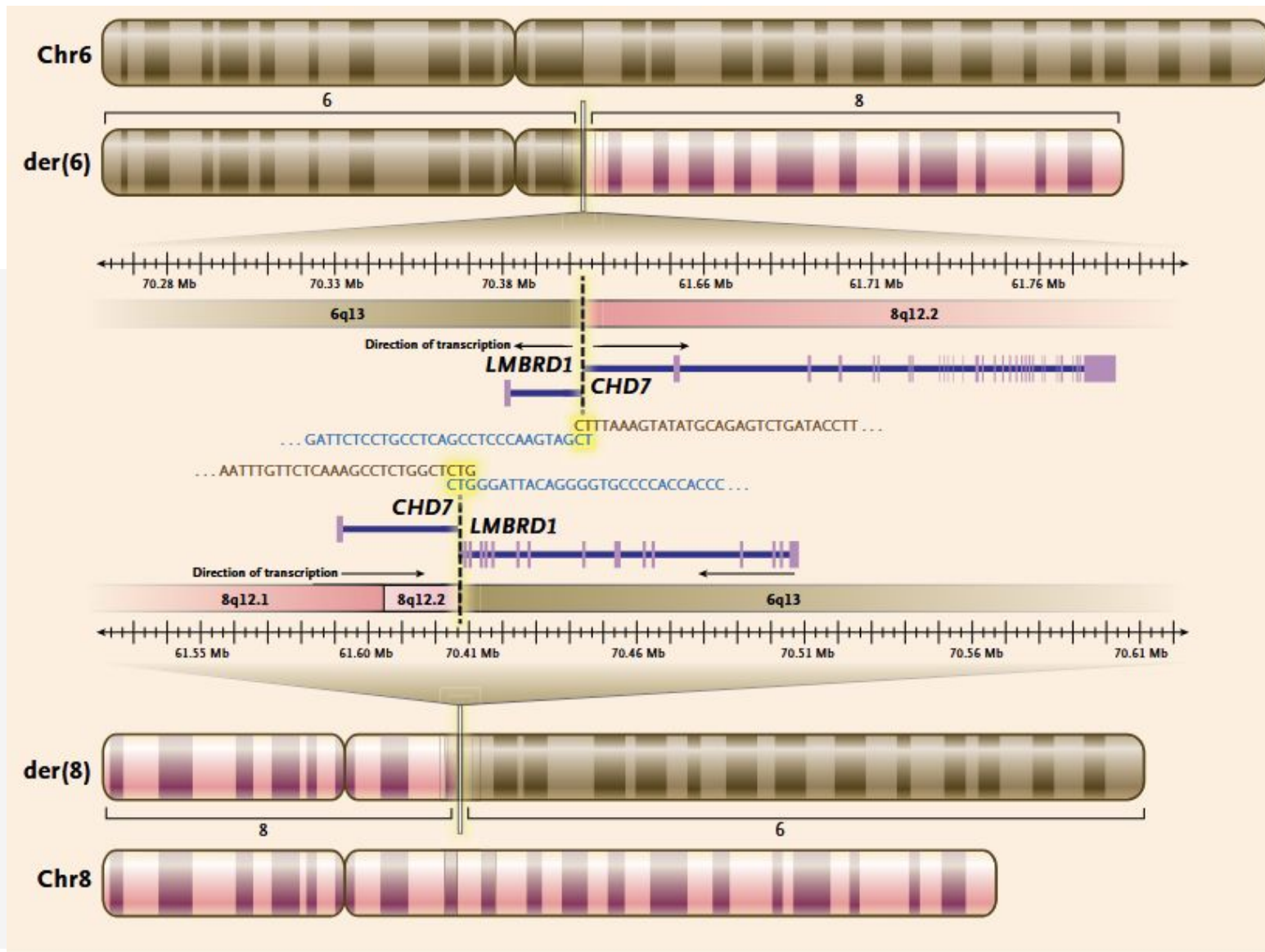


Figure 3. Sequence-Based Delineation of a Balanced De Novo Translocation.

Talkowski, M. E. et al. (2012). Clinical diagnosis by whole-genome sequencing of a prenatal sample. *The New England journal of medicine*, 367(23), 2226–2232.

Hallazgos incidentales

ACMG y CCMG: recomiendan reportar variantes LP o P en genes con **alta penetrancia** que se asocian a **enfermedades moderadas o severas de inicio en la niñez**

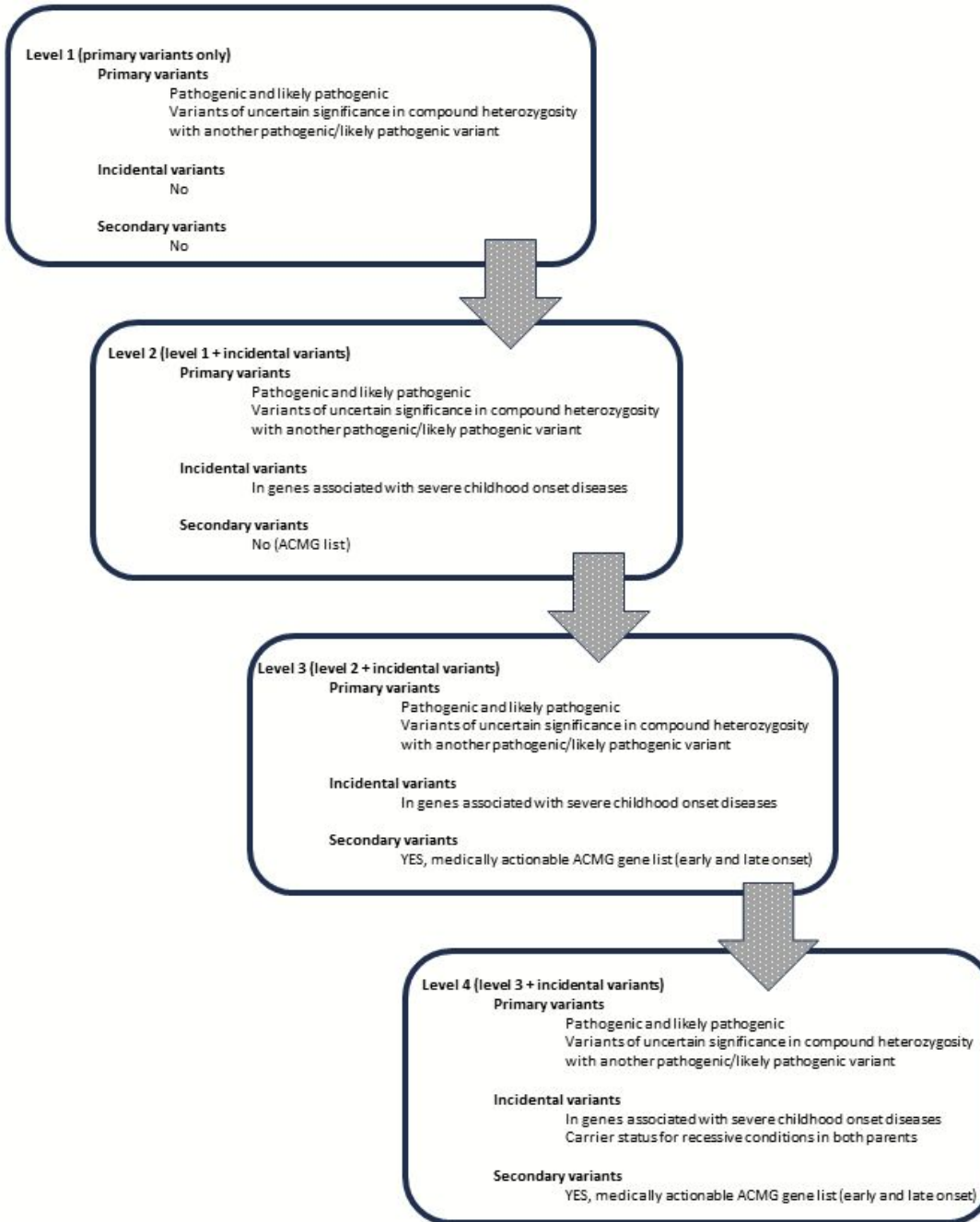
Se recomienda **no reportar estado de portador**

Si se realiza estudio trio, recomiendan cuidado en consejería pre test respecto a informe de hallazgos en progenitores

Hallazgos secundarios

ACMG **sugiere reporte de hallazgos secundarios**
en escenario prenatal

Otros no recomiendan hacerlo (NHS, 2020;
European Society of Human Genetics, 2021;
Canadian College of Medical Geneticists, 2022),
pero dejan a criterio final del clínico



Propuesta de
modelo
escalonado para
reporte de
resultados
prenatales

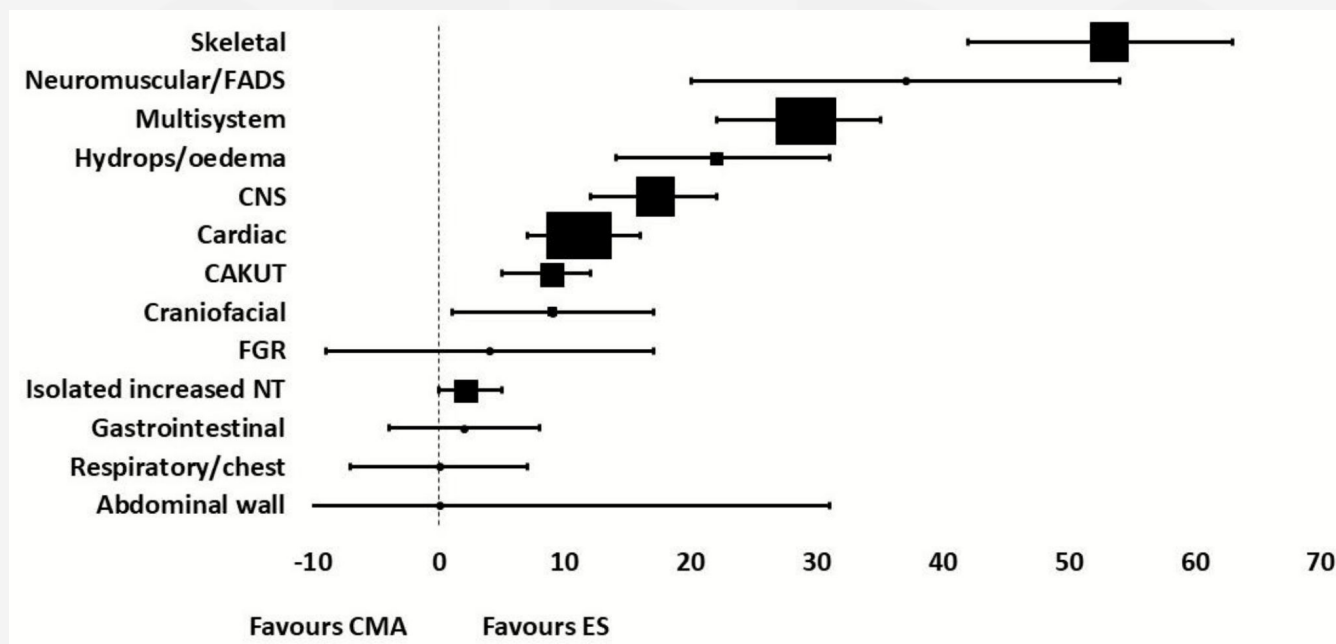
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Rendimientos

WES: rendimiento adicional a cariógrama + CMA es variable según motivo de indicación:

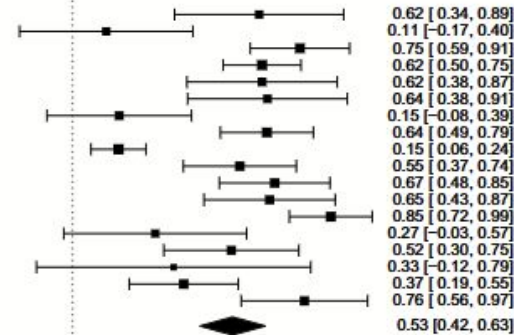


Mellis, R. et al. (2022). Diagnostic yield of exome sequencing for prenatal diagnosis of fetal structural anomalies: A systematic review and meta-analysis. *Prenatal diagnosis*, 42(6), 662–685.

Skeletal

Zhou, X et al, 2018	8	12	66.7
Zhou, J et al, 2021	1	8	12.5
Zhang, X et al, 2021	21	27	77.8
Zhang, L et al, 2021	35	55	63.6
Tang et al, 2021	10	15	66.7
Qi et al, 2020	9	13	69.2
Petrovski et al, 2019	2	12	16.7
Peng et al, 2021	25	38	65.8
Lord et al, 2019	10	65	15.4
Liu et al, 2019	16	28	57.1
Kucinska-Chahwan et al, 2021	18	26	69.2
He et al, 2021	13	19	68.4
Han et al, 2020	23	26	88.5
Fu et al, 2018	3	10	30
Deden et al, 2020	11	20	55
Daum et al, 2019	2	5	40
Chen et al, 2020	11	29	37.9
Chandler et al, 2018	13	16	81.3

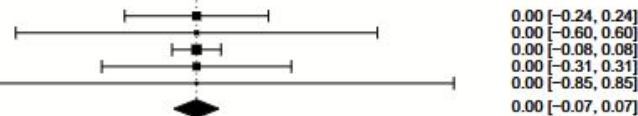
RE Model for Skeletal subgroup



Respiratory

Zhou, J et al, 2021	0	7	0
Petrovski et al, 2019	0	2	0
Lord et al, 2019	0	23	0
Lei et al, 2021	0	5	0
Daum et al, 2019	0	1	0

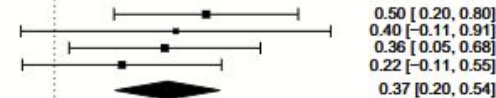
RE Model for Respiratory subgroup



Neuromuscular/FADS

Reischer et al, 2020	6	11	54.5
He et al, 2021	2	4	50
Cao et al, 2021	4	10	40
Boissel et al, 2018	2	8	25

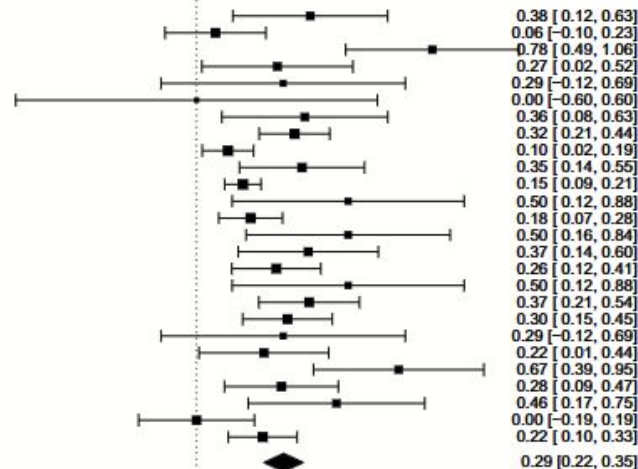
RE Model for Neuromuscular/FADS subgroup



Multisystem

Zhou, X et al, 2021	6	15	40
Zhou, J et al, 2021	1	15	6.7
Zhen et al, 2021	7	8	87.5
Westphal et al, 2019	4	14	28.6
Wagner et al, 2021	2	6	33.3
Sun et al, 2021	0	2	0
Sun et al, 2020	5	13	38.5
Sparks et al, 2020	21	64	32.8
Qiao et al, 2021	6	57	10.5
Qi et al, 2020	8	22	36.4
Lord et al, 2019	22	143	15.4
Liao et al, 2021	4	7	57.1
Li, R et al, 2020	10	55	18.2
Li, L et al, 2020	5	9	55.6
Lei et al, 2021	7	18	38.9
Lei et al, 2020	10	37	27
Heide et al, 2020	4	7	57.1
He et al, 2021	13	34	38.2
Fu et al, 2018	12	39	30.8
Deng et al, 2020	2	6	33.3
Deden et al, 2020	4	17	23.5
De Koning et al, 2021	8	11	72.7
Daum et al, 2019	7	24	29.2
Correa et al, 2021	6	12	50
Chen et al, 2020	0	9	0
Boissel et al, 2018	12	54	22.2

RE Model for Multisystem subgroup

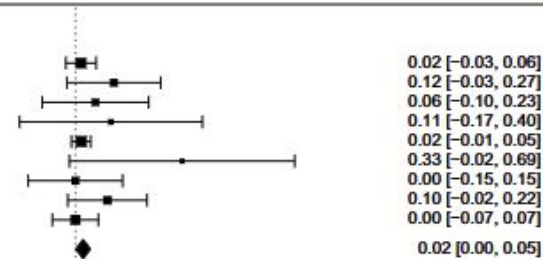


Mellis, R. et al. (2022).
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**Isolated increased NT**

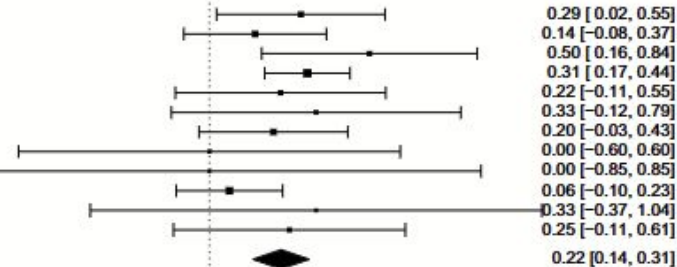
Yang et al, 2020	1	57	1.8
Xue et al, 2020	3	24	12.5
Sparks et al, 2020	1	15	6.7
Qi et al, 2020	1	8	12.5
Mellis et al, 2021	2	111	1.8
Lei et al, 2021	3	8	37.5
Daum et al, 2019	0	12	0
Choy et al, 2019	3	29	10.3
Chen et al, 2020	0	26	0

RE Model for Isolated increased NT subgroup

**Hydrops/Oedema**

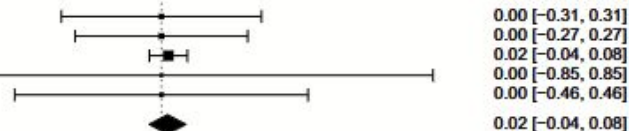
Zhou, X et al, 2021	4	13	30.8
Zhou, J et al, 2021	2	13	15.4
Wagner et al, 2021	5	9	55.6
Sparks et al, 2020	15	48	31.3
Qi et al, 2020	2	8	25
Petrovski et al, 2019	2	5	40
Mone et al, 2021	3	14	21.4
Lei et al, 2021	0	2	0
He et al, 2021	0	1	0
Deng et al, 2020	1	15	6.7
Daum et al, 2019	1	2	50
Correa et al, 2021	2	7	28.6

RE Model for Hydrops/Oedema subgroup

**Gastrointestinal**

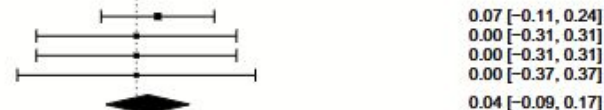
Zhou, J et al, 2021	0	5	0
Petrovski et al, 2019	0	6	0
Lord et al, 2019	1	45	2.2
Lei et al, 2021	0	1	0
Chen et al, 2020	0	3	0

RE Model for GI subgroup

**Fetal growth restriction**

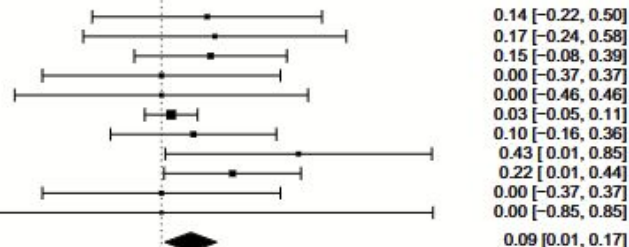
Zhou, J et al, 2021	1	14	7.1
Qi et al, 2020	0	5	0
Petrovski et al, 2019	0	5	0
Lei et al, 2021	0	4	0

RE Model for Fetal growth restriction subgroup

**Craniofacial**

Zhou, J et al, 2021	1	6	16.7
Zhen et al, 2021	1	5	20
Zhang, F et al, 2021	2	12	16.7
Qi et al, 2020	0	4	0
Petrovski et al, 2019	0	3	0
Lord et al, 2019	1	32	3.1
Lei et al, 2021	1	9	11.1
He et al, 2021	3	6	50
Fu et al, 2018	4	17	23.5
Daum et al, 2019	0	4	0
Chen et al, 2020	0	1	0

RE Model for Craniofacial subgroup



Mellis, R. et al. (2022). Diagnostic yield of exome sequencing for prenatal diagnosis of fetal structural anomalies: A systematic review and meta-analysis. *Prenatal diagnosis*, 42(6), 662–685.

CNS

Zhou, J et al, 2021	2	12	16.7
Qi et al, 2020	2	11	18.2
Liao et al, 2021	8	17	47.1
Li, L et al, 2020	3	10	30
Lei et al, 2021	2	11	18.2
Heide et al, 2020	8	55	14.5
He et al, 2021	1	6	16.7
Fu et al, 2018	15	65	23.1
Deden et al, 2020	3	7	42.9
De Koning et al, 2021	2	8	25
Daum et al, 2019	2	15	13.3
Chen et al, 2020	3	12	25
Boissel et al, 2018	5	28	17.9
Baptiste et al, 2022	18	160	11.3

RE Model for CNS subgroup

Cardiac

Zhou, J et al, 2021	3	12	25
Westphal et al, 2019	2	16	12.5
Sun et al, 2021	8	17	47.1
Sun et al, 2020	8	53	15.1
Qiao et al, 2021	18	243	7.4
Qi et al, 2020	1	4	25
Petrovski et al, 2019	1	49	2
Mone et al, 2020	14	122	11.5
Li, R et al, 2020	15	190	7.9
Lei et al, 2021	2	10	20
He et al, 2021	1	12	8.3
Fu et al, 2018	7	34	20.6
Daum et al, 2019	0	2	0
Chen et al, 2020	4	9	44.4

RE Model for Cardiac subgroup

CAKUT

Zhou, X et al, 2020	3	41	7.3
Zhou, J et al, 2021	2	10	20
Qi et al, 2020	3	4	75
Petrovski et al, 2019	1	13	7.7
Lord et al, 2019	0	16	0
Lei et al, 2021	1	8	12.5
Lei et al, 2020	10	126	7.9
He et al, 2021	6	11	54.5
Fu et al, 2018	6	26	23.1
Daum et al, 2019	2	5	40
Chen et al, 2020	1	7	14.3
Boissel et al, 2018	0	11	0

RE Model for CAKUT subgroup

Abdominal wall

Petrovski et al, 2019	0	3	0
Lei et al, 2021	0	2	0
He et al, 2021	0	1	0
Chen et al, 2020	0	1	0

RE Model for Abdominal wall subgroup

RE Model for All Studies

p<0.0001

Heterogeneity: Tau²=0.02, I²=96.4%



0.15 [-0.08, 0.39]
0.17 [-0.09, 0.42]
0.44 [0.20, 0.69]
0.27 [-0.03, 0.57]
0.17 [-0.09, 0.42]
0.14 [0.05, 0.24]
0.14 [-0.22, 0.50]
0.23 [0.12, 0.33]
0.38 [-0.01, 0.76]
0.22 [-0.11, 0.55]
0.12 [-0.07, 0.32]
0.23 [-0.03, 0.49]
0.17 [0.02, 0.32]
0.11 [0.06, 0.16]
0.17 [0.12, 0.22]

0.23 [-0.03, 0.49]
0.12 [-0.07, 0.30]
0.44 [0.20, 0.69]
0.15 [0.05, 0.25]
0.07 [0.04, 0.11]
0.20 [-0.28, 0.68]
0.02 [-0.03, 0.07]
0.11 [0.06, 0.17]
0.08 [0.04, 0.12]
0.18 [-0.09, 0.46]
0.08 [-0.13, 0.28]
0.20 [0.06, 0.34]
0.00 [-0.60, 0.60]
0.40 [0.06, 0.74]
0.11 [0.07, 0.16]

0.07 [-0.02, 0.16]
0.18 [-0.09, 0.46]
0.60 [0.12, 1.08]
0.07 [-0.12, 0.26]
0.00 [-0.11, 0.11]
0.11 [-0.17, 0.40]
0.08 [0.03, 0.13]
0.50 [0.20, 0.80]
0.22 [0.05, 0.39]
0.33 [-0.12, 0.79]
0.12 [-0.19, 0.44]
0.00 [-0.16, 0.16]
0.09 [0.05, 0.12]

0.00 [-0.46, 0.46]
0.00 [-0.60, 0.60]
0.00 [-0.85, 0.85]
0.00 [-0.85, 0.85]
0.00 [-0.31, 0.31]

0.23 [0.19, 0.27]

Mellis, R. et al. (2022).
Diagnostic yield of exome
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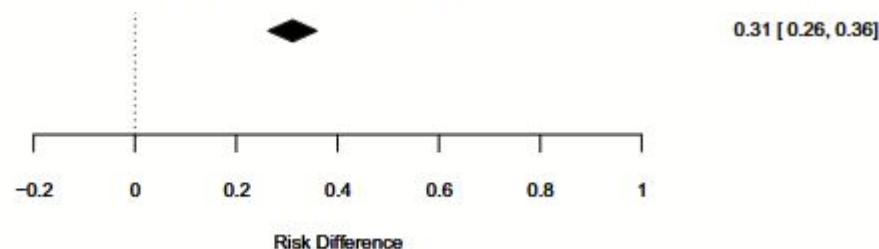
TABLE 3 Pooled effect size for incremental diagnostic yield of ES over CMA in different phenotypic groups

Phenotypic group	Cases analysed	Pooled estimated diagnostic yield [95% CI], <i>p</i> -value
Skeletal	424	53% [42%–63%], <i>p</i> < 0.0001
Neuromuscular/Fetal akinesia deformation sequence (FADS)	33	37% [20%–54%], <i>p</i> < 0.0001
Multisystem	698	29% [22%–35%], <i>p</i> < 0.0001
Hydrops/Oedema	137	22% [14%–31%], <i>p</i> < 0.0001
Central nervous system (CNS)	417	17% [12%–22%], <i>p</i> < 0.0001
Cardiac	773	11% [7%–16%], <i>p</i> < 0.0001
Craniofacial	99	9% [1%–17%], <i>p</i> = 0.02
Congenital anomalies of kidneys and urinary tract (CAKUT)	278	9% [5%–12%], <i>p</i> < 0.0001
Fetal growth restriction	28	4% [–9 to 17%], <i>p</i> = 0.59
Isolated increased nuchal translucency (NT)	290	2% [0%–5%], <i>p</i> = 0.04
Gastrointestinal	60	2% [–4 to 8%], <i>p</i> = 0.5
Respiratory/Chest	38	0 [–7 to 7%], <i>p</i> = 1
Abdominal wall	7	0 [–31% to 31%], <i>p</i> = 1

Note: Phenotypic groups refer to fetuses with one or more anomalies in a single body system. Fetuses with anomalies in more than one system are classified as 'Multisystem'. The 'isolated increased NT' group contains a mixture of (i) cases with isolated increased NT at presentation where it was unspecified whether additional anomalies developed later in pregnancy, and (ii) cases where isolated increased NT remained isolated throughout pregnancy. The bold and underlined formatting was just for emphasis in defining the two sub-groups.

Abbreviations: CMA, chromosomal microarray; ES, exome sequencing.

RE Model
p < 0.0001
Heterogeneity: $\tau^2 = 0.03$, $I^2 = 94.0\%$



Riesgo agrupado:
0.31

WGS: rendimiento

Summary of fetal exome sequencing publications with >5 fetuses included

First author	Number of cases	Cohort description	Proband vs. trio	Pathogenic variant	Likely pathogenic variant
Yang et al., 2014	11	Terminated anomalous fetus	Trio	6 of 11 (54%)	—
Carss et al., 2014	30	Prenatal sonographic anomalies	Trio	3 of 30 (10%)	5 of 30 (16.7%)
Drury et al., 2015	24	Prenatal sonographic anomalies including NT $\geq$ 3.5mm	14 Proband10 Trio	5 of 24 (20.8%)	1 of 24 (4.2%)
Alamillo et al., 2015	7	Multiple sonographic anomalies termination or demise	Trio	3 of 7 (42.9%)	1 of 7 (14.3%)
Pangalos et al., 2017	14	Prenatal sonographic anomalies	Proband only	6 of 14 (42.9%)	—
Yates et al., 2017	84	Demise or termination	33 Proband/duo51 Trio/quad	17 of 84 (20%)	38 of 84 (45%)
Vora et al., 2017	15	Multiple sonographic anomalies	Trio	7 of 15 (46.7%)	1 of 15 (6.7%)
Fu et al., 2018	196	Prenatal sonographic anomalies	34 Proband13 Trio	47 of 196 (24%)	25 of 196 (12.8%)

WGS: rendimiento

Table 2 Incremental yield of whole-genome sequencing compared with quantitative fluorescence polymerase chain reaction/ chromosomal microarray analysis with resolution of 50 kb

<i>Anomaly group</i>	<i>Incremental yield (%)</i>	<i>Heterogeneity (%)</i>
All		
All cases	26 (18–36)	86
Prenatal	16 (9–24)	85
Postnatal	39 (27–51)	53
Multisystem		
All cases	30 (19–43)	65
Prenatal	23 (12–36)	70
Postnatal	43 (27–59)	21
R21 selected		
All cases	32 (22–42)	70
Prenatal	22 (14–31)	65
Postnatal	48 (37–60)	0

Shreeve, N. et al. (2024). Incremental yield of whole-genome sequencing over chromosomal microarray analysis and exome sequencing for congenital anomalies in prenatal period and infancy: systematic review and meta-analysis. *Ultrasound in obstetrics & gynecology : the official journal of the International Society of Ultrasound in Obstetrics and Gynecology*, 63(1), 15–23.

Overview

Rapid prenatal exome sequencing (**R21** in the [National Genomic Test Directory](#)) is undertaken for at-risk pregnancies in which a genomic diagnosis would guide management of the fetus. At present, **R21** testing may only be requested by a clinical geneticist, usually following multidisciplinary team discussion with the fetal medicine team. Testing requires invasive prenatal sampling and involves [exome sequencing](#) of a nationally approved panel of genes known to cause prenatal genetic disorders.

What are the R21 testing eligibility criteria?

R21 testing can be requested when:

- a fetus presents with multiple anomalies affecting multiple systems;
and/or
- the presentation is suggestive of an underlying monogenic disorder;
and
- a molecular diagnosis may influence the management of the pregnancy or the baby in the immediate neonatal period.

Some examples of presentations for which **R21** testing may be indicated include a fetus with:

- multiple anomalies in a number of different systems;
- a suspected skeletal dysplasia where intrauterine growth restriction has been excluded;
- large echogenic kidneys and a normal bladder;
- major central nervous system anomalies in which a neural tube defect has been excluded;
- multiple contractures (excluding bilateral isolated talipes);
- a [nuchal translucency](#) greater than 6.5 millimetres **and** at least one additional anomaly and in which the array comparative genomic hybridisation (CGH) is normal; and
- non-immune fetal hydrops (fluid/oedema in at least two compartments detected at or after routine second trimester scan) and a normal array CGH.

Please also refer to the test directory for the complete lists of eligibility and exclusion criteria.

Exome sequencing in every pregnancy? Results of trio exome sequencing in structurally normal fetuses

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- Análisis retrospectivo, centro único. Fetos estructuralmente normales
- Exoma clínico
- 1825 exomas, **1020 casos de bajo riesgo** con 28 fetos (**2.7%**) con variantes potencialmente patogénicas, con 64% de novo dominantes
- Terminación voluntaria del embarazo en 13 casos (1.2%), incluyendo 4/7 VUS

Rendimientos - Obito

Quinlan-Jones (2019): 27 fetos abortados estructuralmente anormales con CMA y QF-PCR normal

- Rendimiento de exoma tríó **37%**
- 4 variantes de novo, 6 restantes heredadas AR o ligadas a X

Yates (2017): 84 fetos, exoma tríó

- **20% positivos**, 45% posibles, 9% gen candidato
- 24% positivos en WES tríó, 14% en WES solo

Otros



> J Perinat Med. 2022 Dec 13;51(6):769-774. doi: 10.1515/jpm-2022-0504. Print 2023 Jul 26.

Rapid diagnosis of intra-amniotic infection using nanopore-based sequencing

Piya Chaemsaithong^{1 2 3 4}, Roberto Romero^{3 4 5 6 7 8}, Pisut Pongchaikul^{9 10 11},

Conclusiones

- NGS presenta **rendimiento variable según la indicación clínica**, y requiere criterio según recursos disponibles para indicar
- WGS promete gran rendimiento. Sin embargo, **se necesita evidencia comparativa adicional** para poder sugerir por sobre exoma